

Microbiome, Immune function, and murine pulmonary disease

CURES Symposium:

Addressing the Asthma and Allergy Epidemics

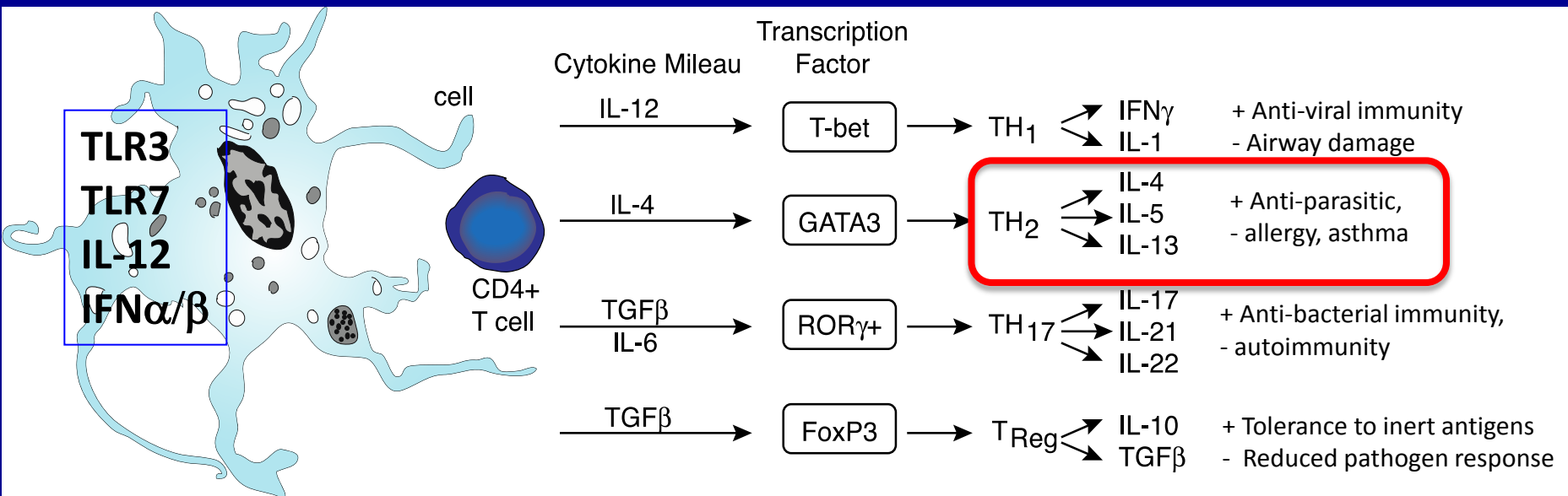
October 7, 2015

Nick Lukacs, PhD

Godfrey Dorr Stobbe Professor of Pathology

University of Michigan Medical School

T cell maturation and differentiation depends upon Immune environments

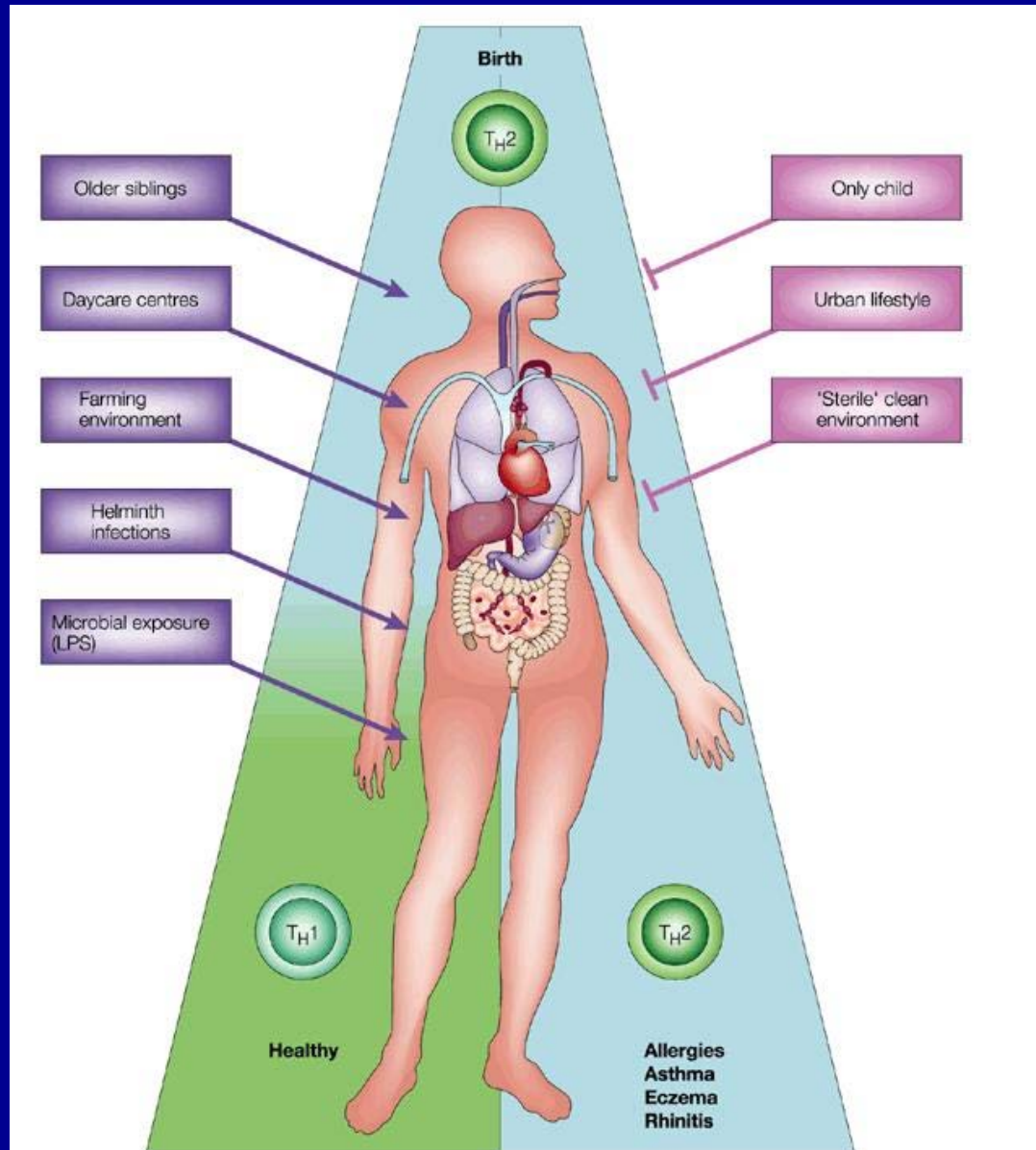


IL-4- IgE → Mast cell activation

IL-5- Eosinophilia → airway damage and fibrosis

IL-13- Goblet cell metaplasia → mucus and airway obstruction

The hygiene hypothesis





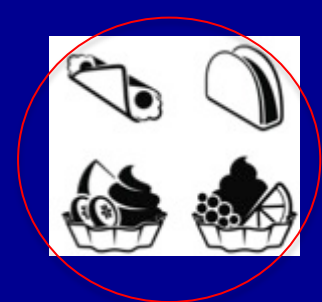
Microbial Exposures



Host genetics

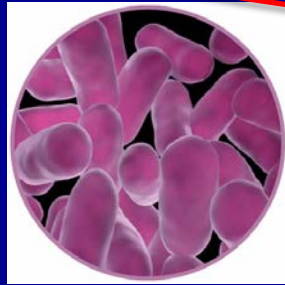


Pharmaceuticals

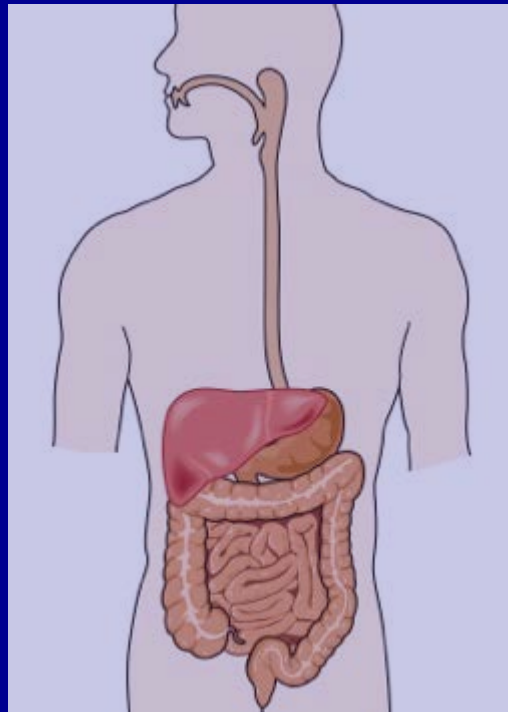


Food

Factors that affect the gut microbiome



Commensal bacteria



Pathobionts

Protection

- Mucosal barrier function
- Treg cell development

Development and modulation of the host immune responses

Metabolism

The resulting overgrowth of the pathobiont may cause inflammation and bleeding of the lining of the colon.

Obesity
Cancer

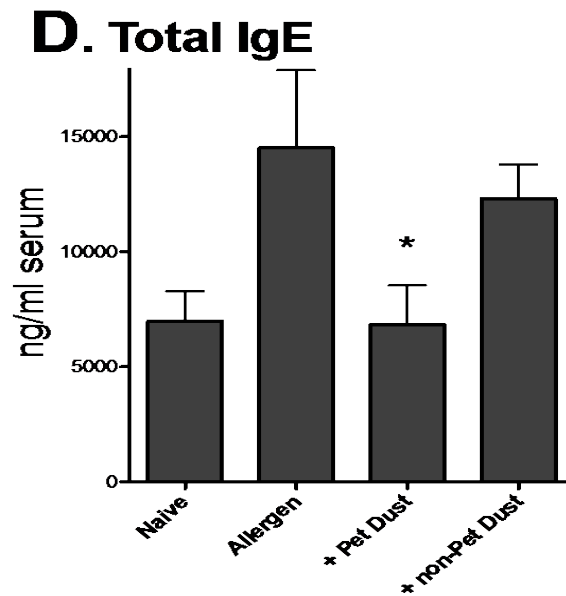
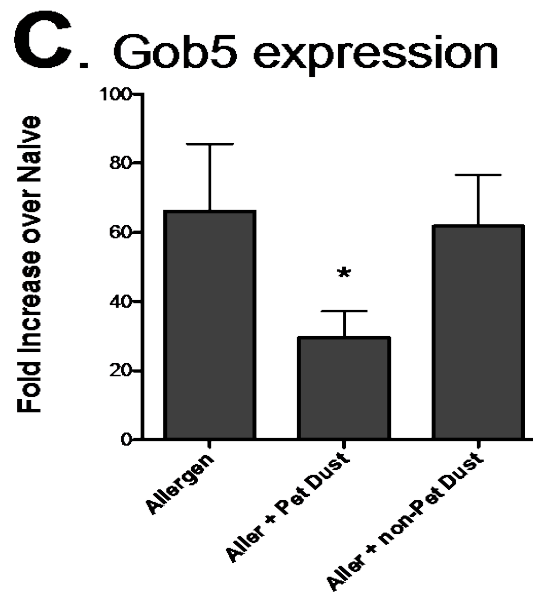
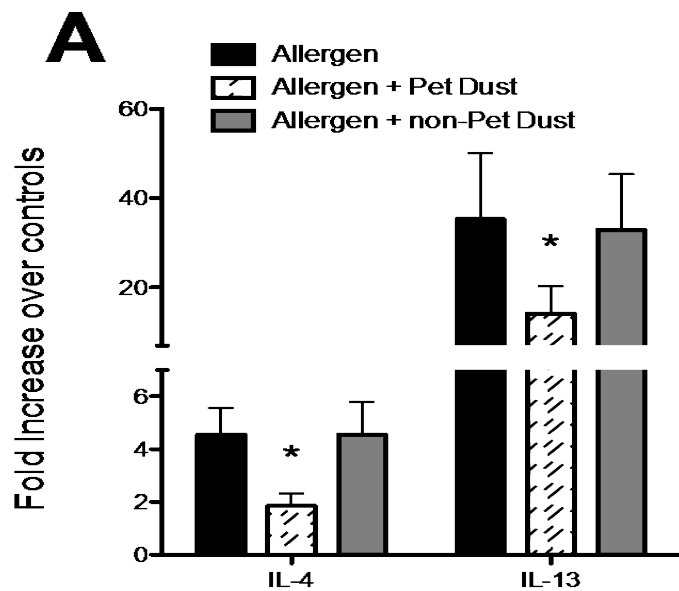
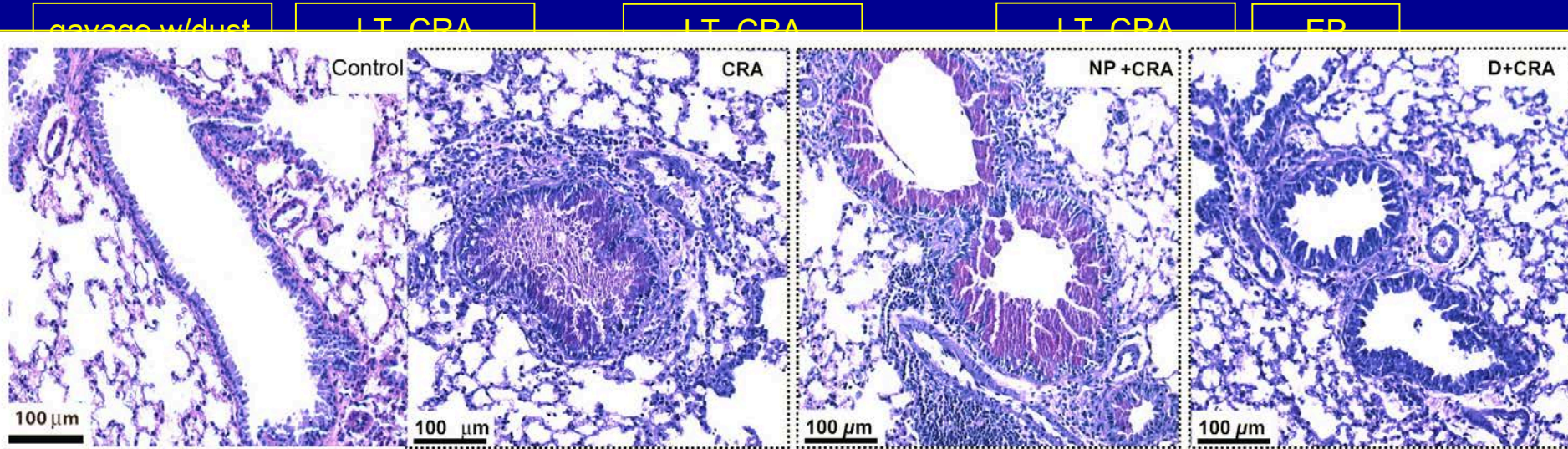
Infant's Environment shapes the Microbiome

- Natural vs. Cesarean section birth
- Bottle vs. Breast feeding
- Timing and type of solid food introduction
- Antibiotic use
- Vitamin and nutrition
- Household exposure- high % of early life

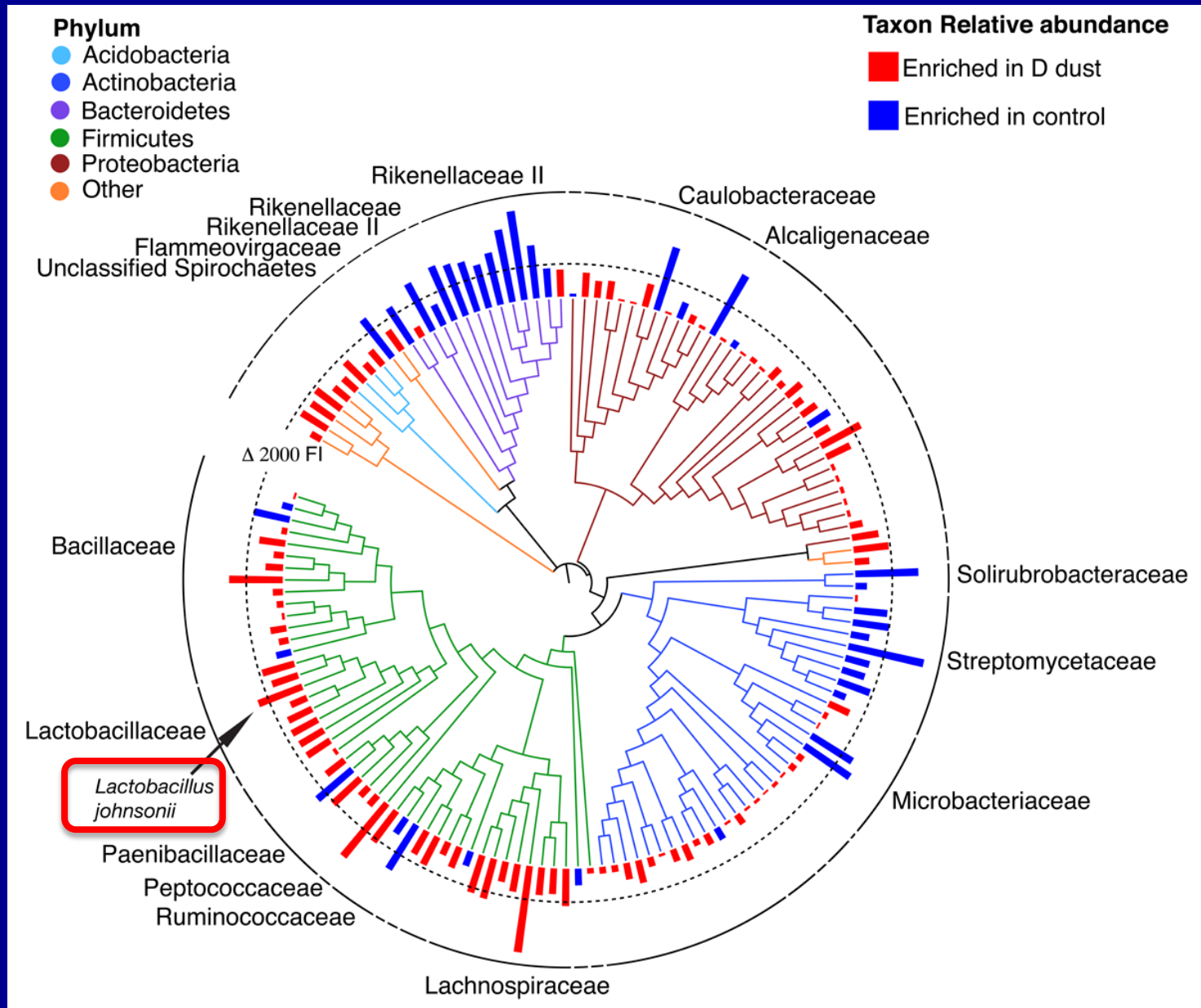
Man's best friend? The effect of pet ownership on house dust microbial communities.
-Increase in bacterial diversity and a decrease in fungal species

Fujimura KE, Johnson CC, Ownby DR, Cox MJ, Brodie EL, Havstad SL, Zoratti EM, Woodcroft KJ, Bobbitt KR, Wegienka G, Boushey HA, Lynch SV. JACI 126:410.

Dust-exposed mice have a modified response in cockroach allergen (CRA) model



Bacterial diversity in Pet dust supplemented animals

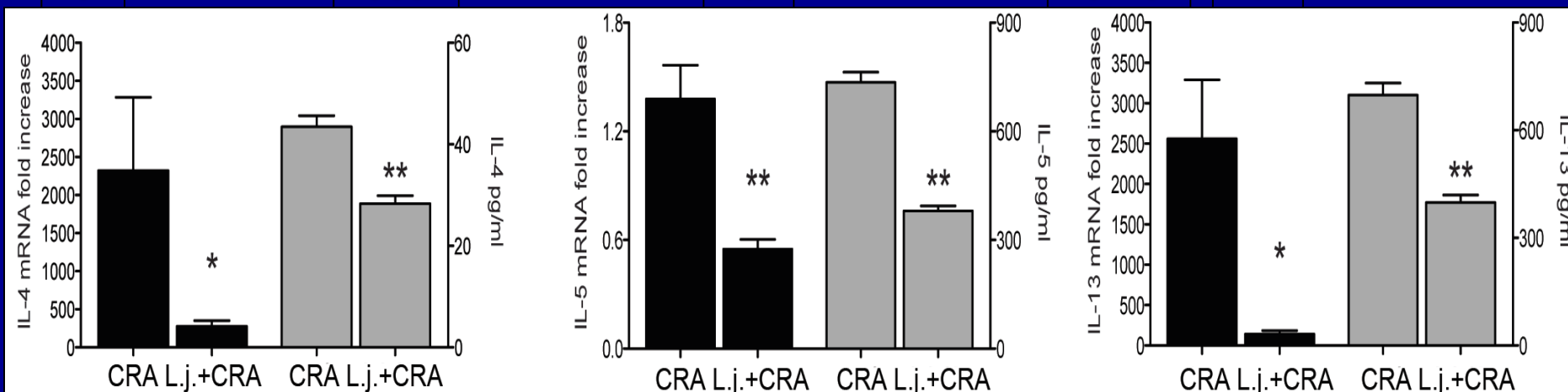
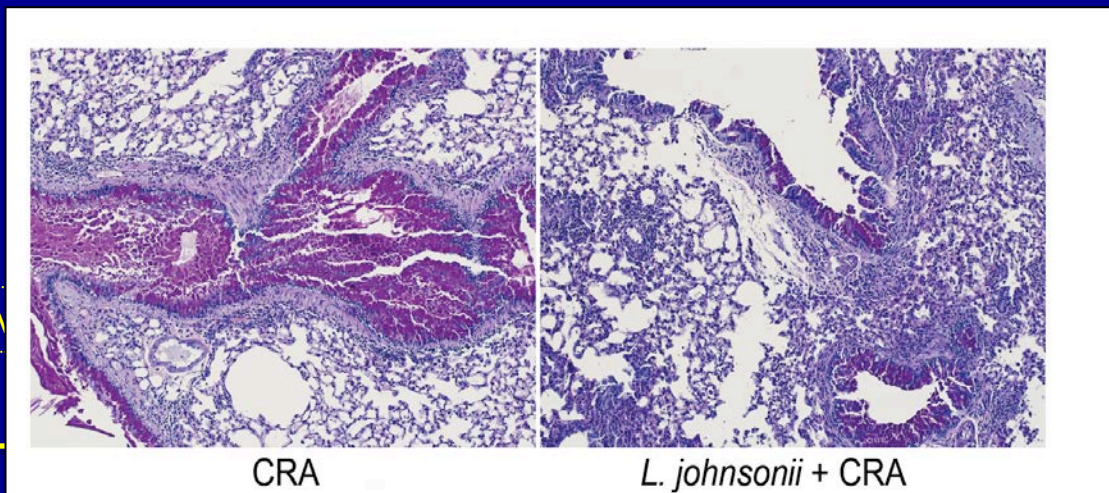
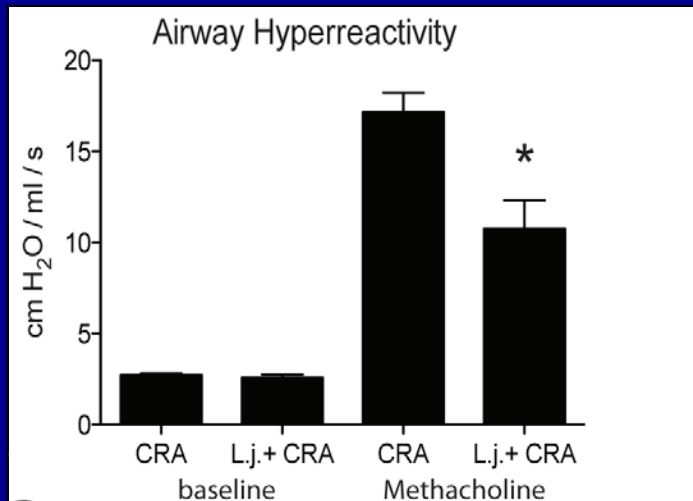


Lactobacillus Supplementation

- Colonic contents of 4 mice
- *Lactobacillus* isolation media
- Sequenced 6 isolates per mouse
- Twenty one isolates yielded high quality full length 16S rRNA sequence— **All were *L. johnsonii***
- 99% coverage and 99% homology to expected *Lactobacillus* species
- Batch culture
- Standardized (1×10^7 CFU) supplements in glycerol



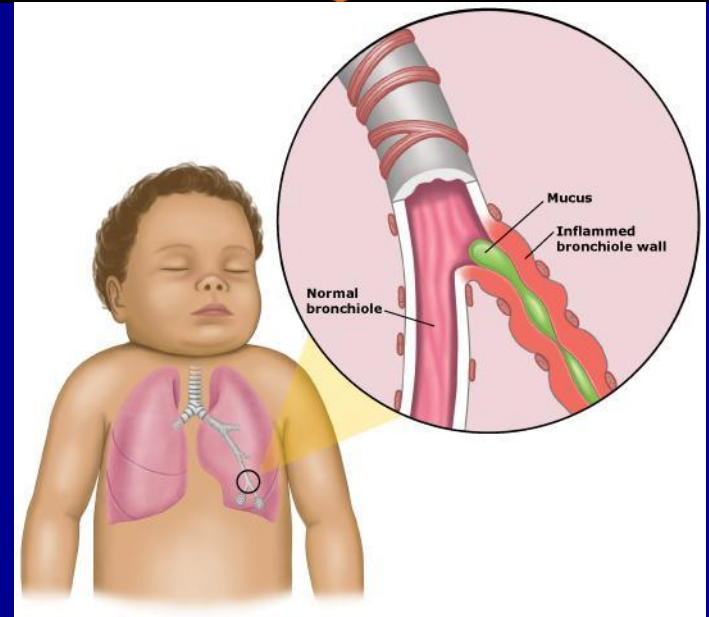
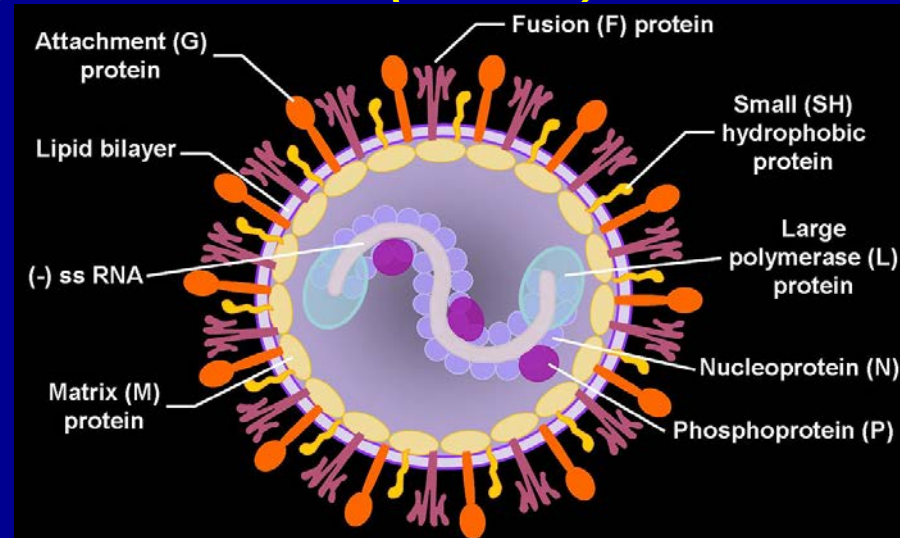
Lactobacillus johnsonii supplementation protects asthmatic mice



Viral infections during infancy -> Asthma?

Respiratory Syncytial Virus (RSV)

- ❑ Leading cause of respiratory illness and hospitalization in infants
- ❑ Airway epithelial damage
- ❑ Leads to long-term Respiratory disease
- ❑ Goblet cell hypertrophy, mucus hypersecretion;
- ❑ Th2 and Th17 cytokine production;
- ❑ Associated with increased Asthma
- ❑ During RSV infection-Tregs control the magnitude of cellular immune responses. (Brincks EL, J. Immunology, 2013)



L. johnsonii
Supplementation



0

RSV
 10^5
PFU



8

L. johnsonii
Supplementation



9



12

Histology
AHR remodeling
Ceca



16

Time (day)

Fig 1. Mice treatment over course of the experiment

-Viable vs heat killed bacteria

1 x 10^7 CFU daily supplement – 7 days

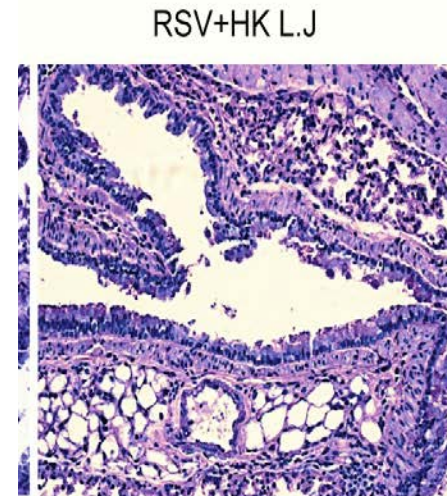
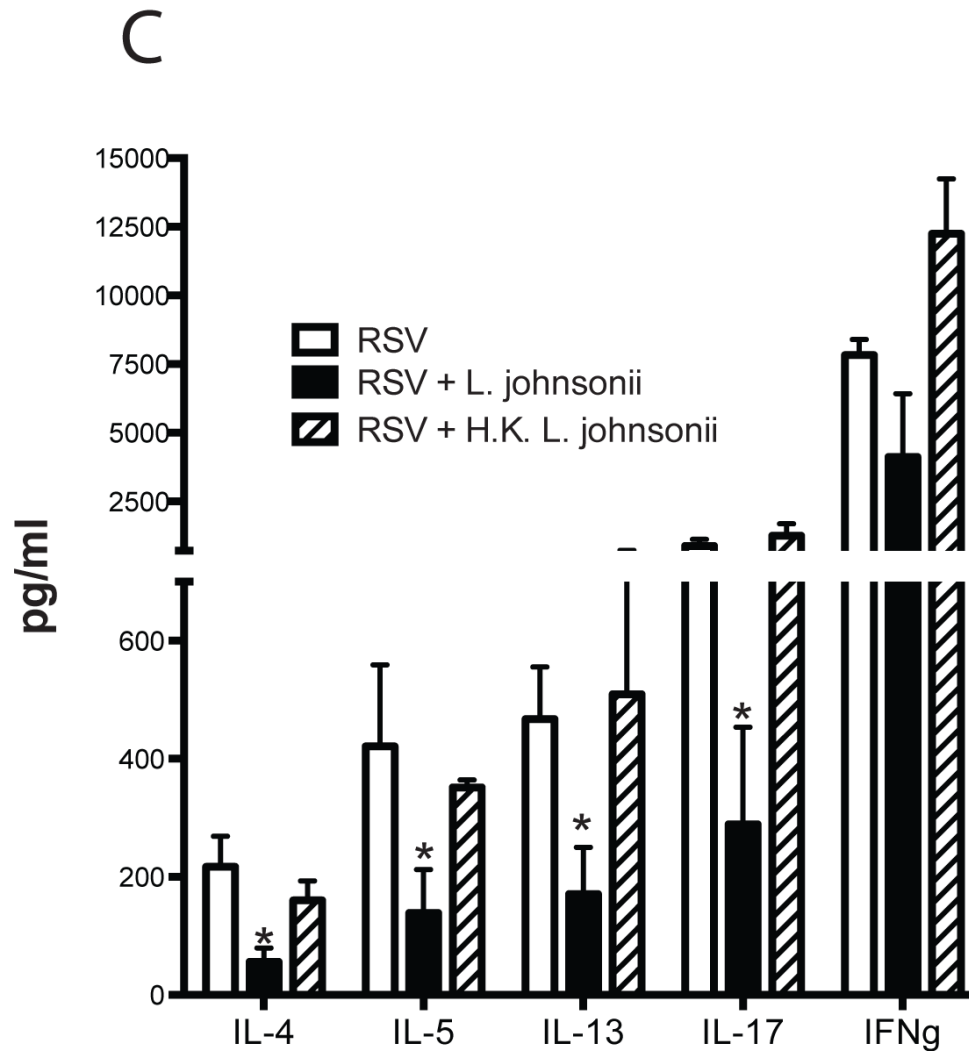
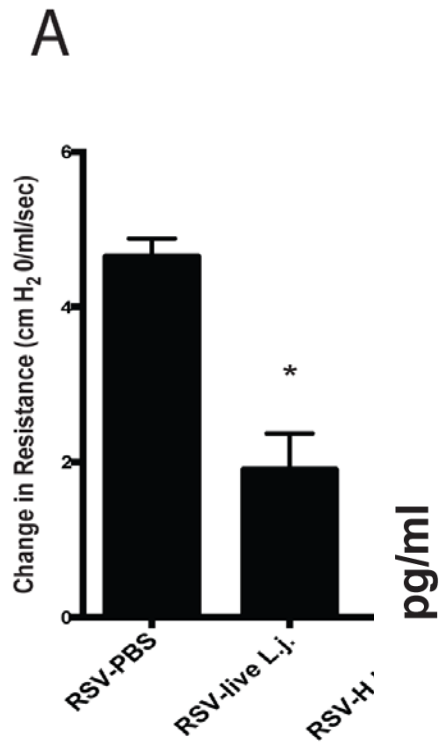
-RSV (line 19) infection on day 8 of treatment protocol

- Outcome measurements

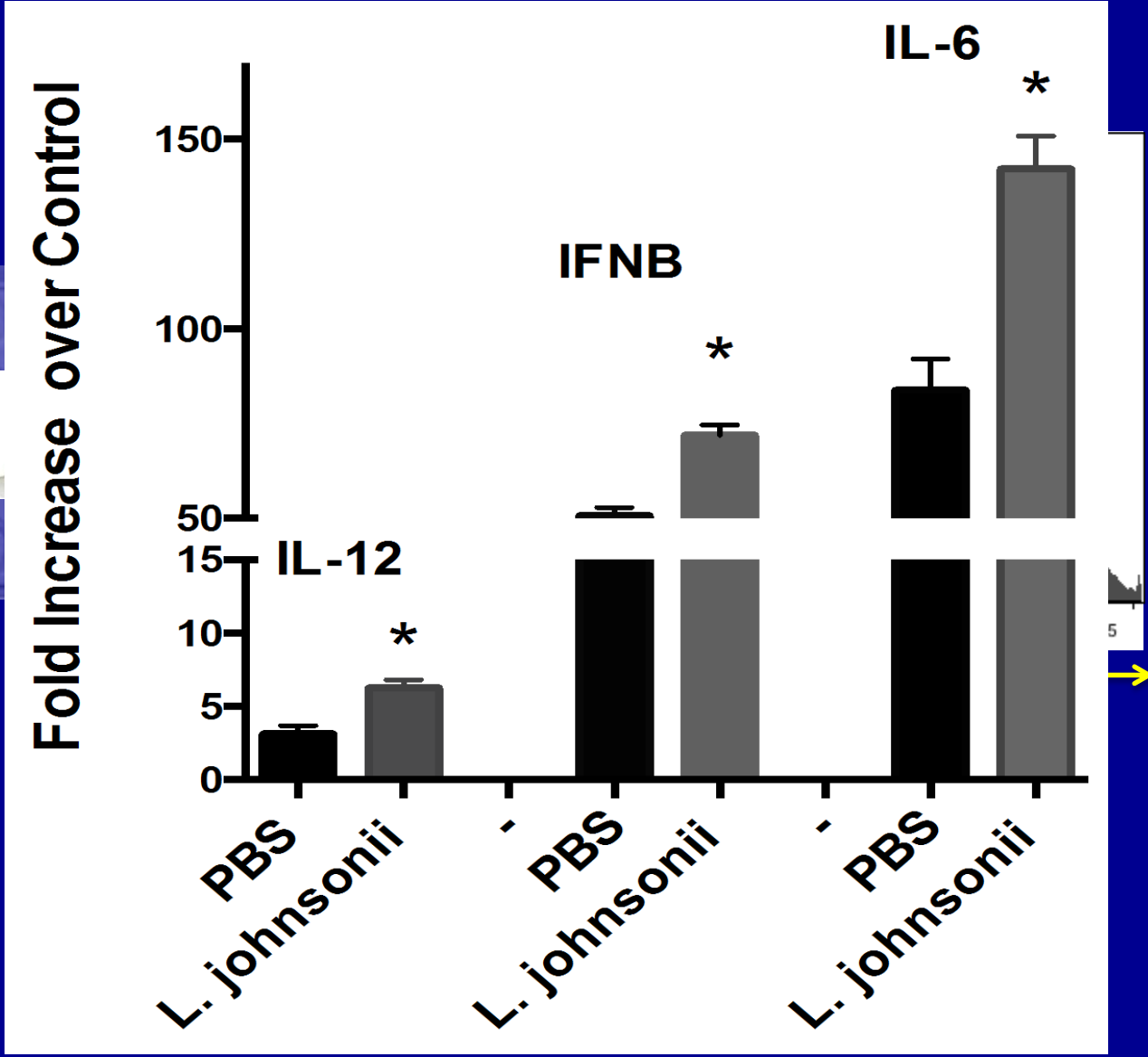
1. Airway responses – histology

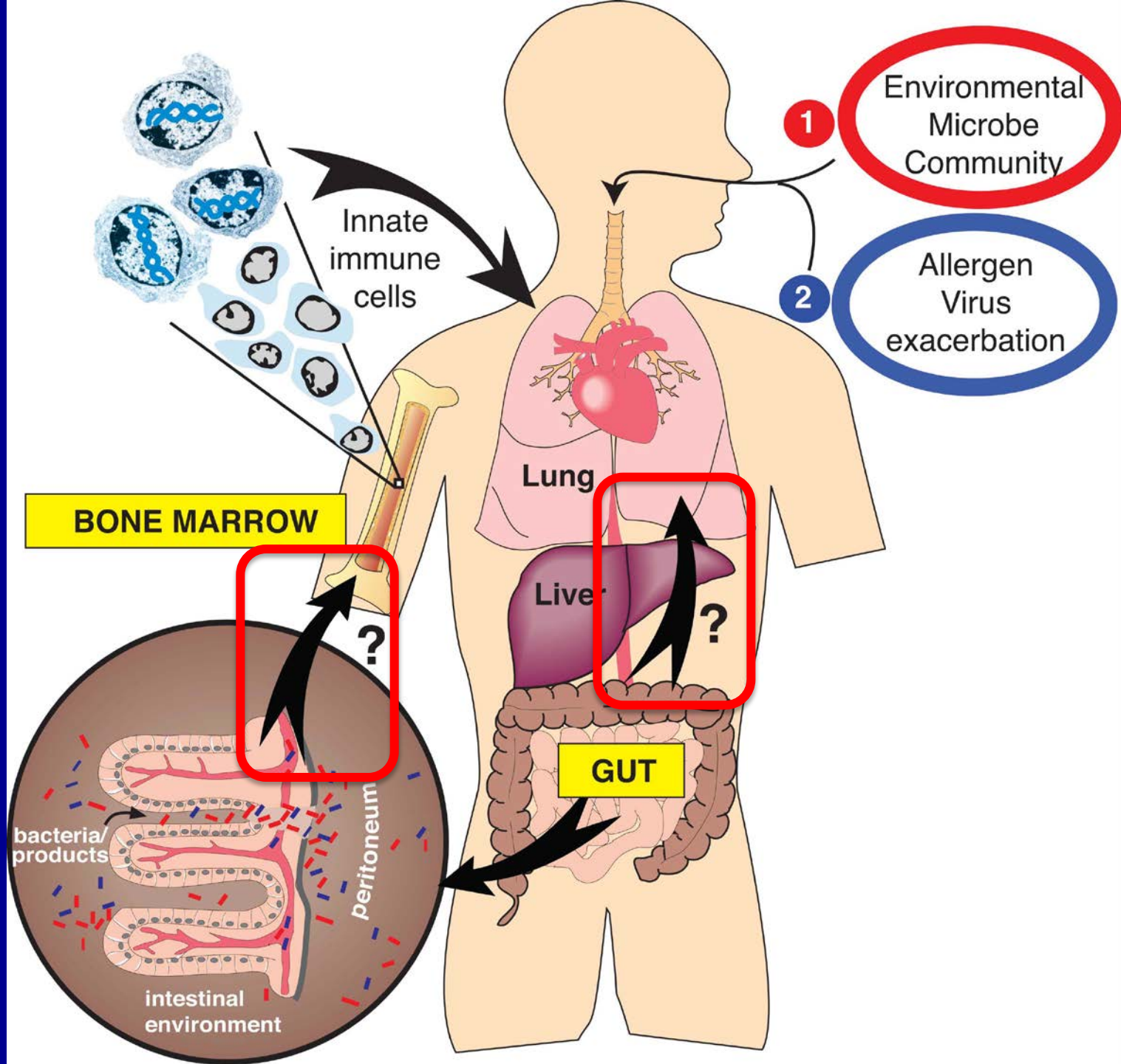
2. Immune responses – Th2, IFN γ , muc5AC Gob5

L. Johnsonii supplementation alters RSV-induced pathophysiology



Bone marrow DC are altered in *L. johnsonii* exposed animals in Response to RSV





1 Environmental Microbe Community

2 Allergen Virus exacerbation

BONE MARROW

GUT

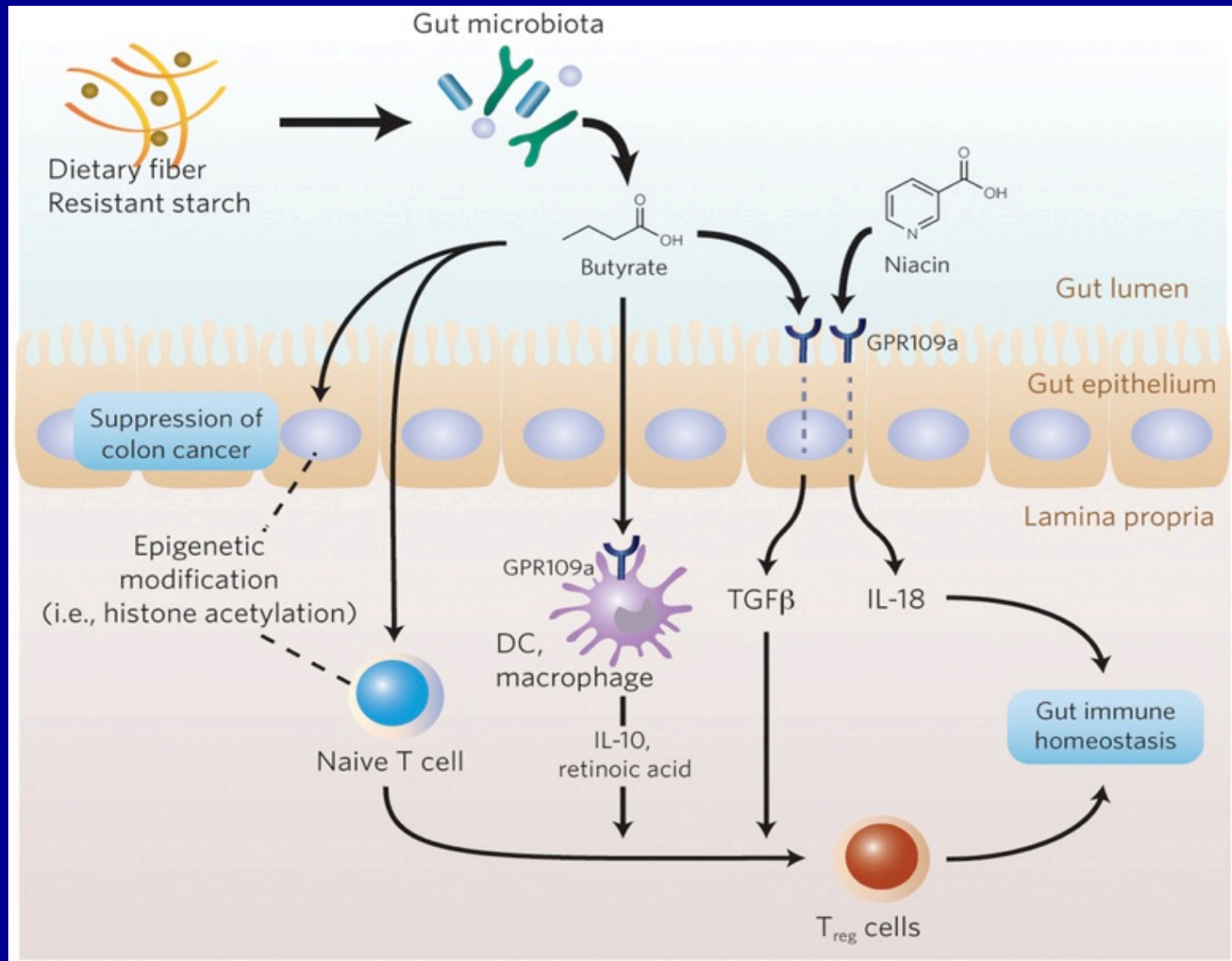
intestinal environment
peritoneum
bacteria/products

Innate immune cells

Lung

Liver

Microbiome can determine metabolite production available to alter immune function



Sub Pathway	Biochemical Name	PBS	L. johnsonii
Leucine, Isoleucine and Valine Metabolism	alpha-hydroxyisovalerate	1.0314	1.2844
	isoleucine	0.95	1.3852
	3-methyl-2-oxovalerate	0.7178	1.8789
	2-hydroxy-3-methylvalerate	1.1784	1.6134
	valine	0.9468	1.3642
	3-methyl-2-oxobutyrate	0.8829	1.5879
	3-hydroxyisobutyrate	1.0203	1.8043
Long chain fatty acids	alpha-hydroxyisocaproate	0.7684	1.8758
	myristoleate (14:1n5)	1.0156	1.6242
	palmitate (16:0)	0.8279	1.2426
	palmitoleate (16:1n7)	0.9297	1.9098
	10-heptadecenoate (17:1n7)	0.9143	1.6254
	oleate (18:1n9)	0.9946	1.5634
	cis-vaccenate (18:1n7)	1.1388	1.6774
Polyunsaturated Fatty Acid (n3 and n6)	10-nonadecenoate (19:1n9)	0.9574	1.7259
	stearidonate (18:4n3)	0.9092	2.0629
	eicosapentaenoate (EPA; 20:5n3)	0.9697	1.6829
	docosapentaenoate (n3 DPA; 22:5n3)	1.0019	2.1902
	docosahexaenoate (DHA; 22:6n3)	1.0062	1.4737
	linolenate [alpha or gamma; (18:3n3 or 6)]	0.9729	1.8151
	docosapentaenoate (n6 DPA; 22:5n6)	0.8102	1.527
Fatty acid Metabolism	dihomo-linoleate (20:2n6)	0.8467	1.6506
	mead acid (20:3n9)	0.8509	2.1137
	myristoylcarnitine	0.929	1.8567
	palmitoylcarnitine	0.8664	1.5805
	stearoylcarnitine	1.0211	2.2988
	oleoylcarnitine	1.0648	1.5524
	myristoleoylcarnitine*	1.0617	1.6484
Fatty Acid, Monohydroxy	2-hydroxyoctanoate	0.7521	1.6925
	2-hydroxydecanoate	0.6441	2.0658
	3-hydroxyoctanoate	1.0549	1.9832
	3-hydroxydecanoate	1.1702	2.1847
	3-hydroxylaurate	1.2756	1.9255
	3-hydroxymyristate	1.072	1.6838
	5-hydroxyhexanoate	1.624	0.9847
Lysolipids	13-HODE + 9-HODE	0.5303	1.2566
	2-palmitoylglycerophosphocholine	0.9996	1.5364
	1-palmitoleoylglycerophosphocholine (16:1)*	0.8732	1.9251
	1-stearoylglycerophosphocholine (18:0)	0.8448	1.5755
	2-stearoylglycerophosphocholine*	1.1226	1.4972
	1-oleoylglycerophosphocholine (18:1)	0.8709	1.5372
	1-linoleoylglycerophosphocholine (18:2n6)	1.0147	1.5413
	1-linolenoylglycerophosphocholine (18:3n3)*	1.0223	2.2415
	1-arachidonoylglycerophosphocholine (20:4n6)*	0.9596	1.4044
	1-palmitoylplasmenyethanolamine*	0.7613	0.8295
	1-oleoylglycerophosphoethanolamine	0.7084	1.4933
	1-linoleoylglycerophosphoethanolamine*	0.7802	1.7731
	1-arachidonoylglycerophosphoethanolamine*	0.8629	1.6604
	1-linoleoylglycerophosphoinositol*	0.8457	1.4562
	1-arachidonoylglycerophosphoinositol*	0.9122	1.2935
	palmitoyl-linoleoyl-glycerophosphocholine (2)*	0.9206	1.2143
	palmitoyl-palmitoyl-glycerophosphocholine (1)*	0.5473	0.9566
palmitoyl-palmitoyl-glycerophosphocholine (2)*	0.4736	0.7721	
stearoyl-linoleoyl-glycerophosphocholine (1)*	0.9828	1.2829	
Monoacylglycerol	1-palmitoylglycerol (1-monopalmitin)	1.2643	1.6405
	1-oleoylglycerol (1-monolein)	0.5198	3.048
	1-linoleoylglycerol (1-monolinolein)	1.0244	2.235
	2-linoleoylglycerol (2-monolinolein)	0.8505	2.9696
	1-docosahexaenoylglycerol	1.462	1.8398
	1-dihomo-linolenylglycerol (alpha, gamma)	1.3659	1.7462
Steroid	2-docosahexaenoylglycerol*	1.8316	2.815
	corticosterone	1.2699	0.5477
	11-dehydrocorticosterone	1.3886	0.3968
Primary Bile Acid Metabolism	cholate	4.5401	18.3219
	taurocholate	0.6944	5.8782
	taurochenodeoxycholate	0.4883	6.1091
	beta-muricholate	1.8429	4.2474
	tauro-beta-muricholate	0.846	6.9928
Secondary Bile Acid Metabolism	deoxycholate	0.877	3.0533
	taurodeoxycholate	0.5989	2.3552
	taurothiocholate 3-sulfate	0.9432	3.1577
	taurohyodeoxycholic acid	0.771	5.2808
	7-ketodeoxycholate	1.9761	19.3096

Upregulation of plasma metabolites in animals supplemented with *L. johnsonii*

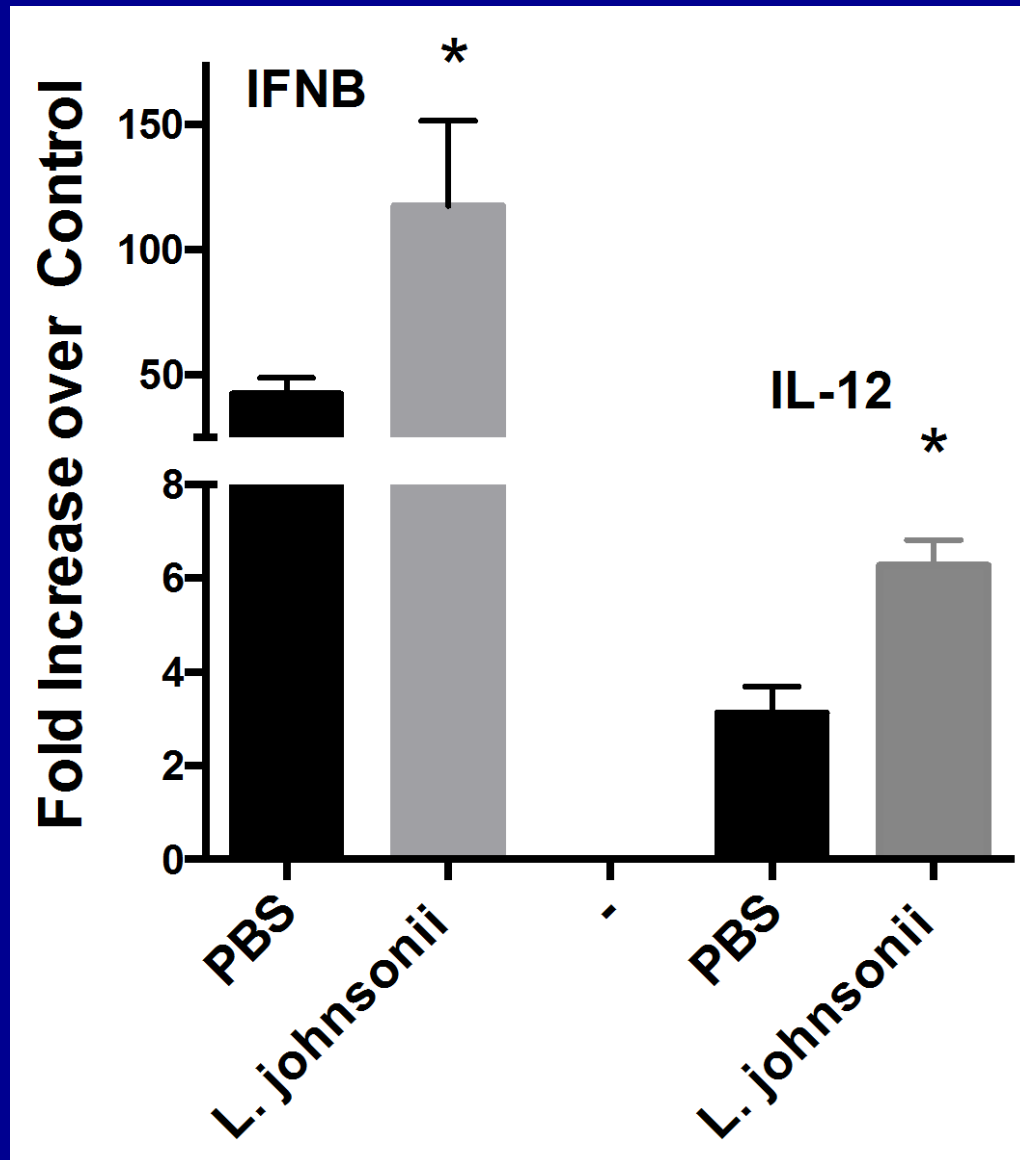
- Animals were supplemented with 1 X 10⁷ cfu of *L. johnsonii* by oral gavage for 7 days and plasma metabolite levels were assessed
- Supplemented animals were then infected with RSV. After 2 days of RSV infection plasma from sacrificed animals (5/group) was harvested and the metabolite levels compared to supplemented mice at day 0 prior to infection.

Red – significantly upregulated

Green- significantly downregulated

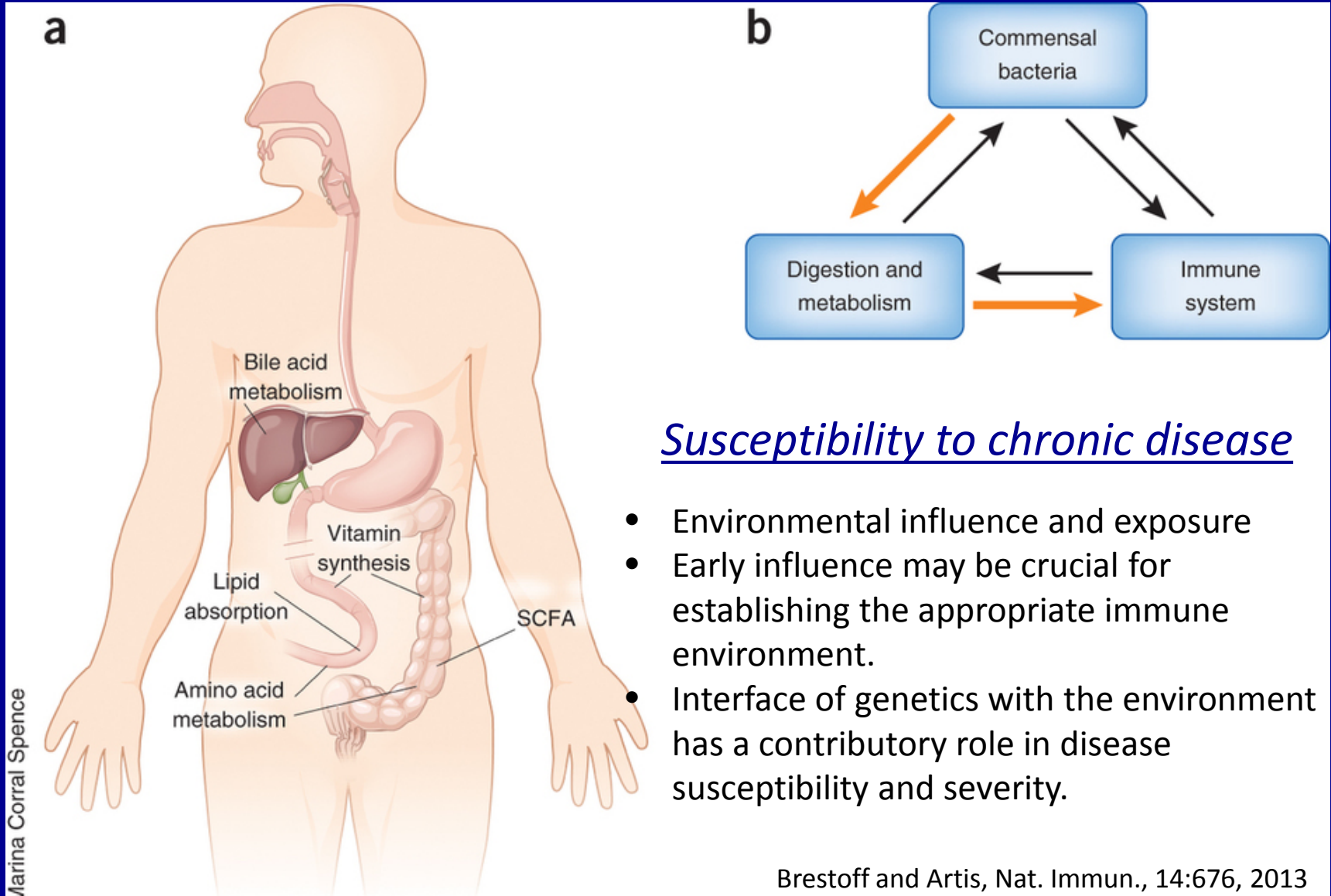
Sub Pathway	Biochemical Name	PBS	L. johnsonii
Leucine, Isoleucine and Valine Metabolism	alpha-hydroxyisovalerate	1.0314	1.2644
	isoleucine	0.95	1.3852
	3-methyl-2-oxovalerate	0.7178	1.8789
	2-hydroxy-3-methylvalerate	1.1784	1.6114
	valine	0.9468	1.3642
	3-methyl-2-oxobutyrate	0.8829	1.5879
	3-hydroxyisobutyrate	1.0203	1.8043
	alpha-hydroxyisocaproate	0.7684	1.8758
Long chain fatty acids	myristoleate (14:1n5)	1.0156	1.6242
	palmitate (16:0)	0.8279	1.2426
	palmitoleate (16:1n7)	0.9297	1.9098
	10-heptadecenoate (17:1n7)	0.9143	1.6254
	oleate (18:1n9)	0.9946	1.5634
	cis-vaccenate (18:1n7)	1.1388	1.6774
	10-nonadecenoate (19:1n9)	0.9574	1.7259
Polyunsaturated Fatty Acid (n3 and n6)	stearidonate (18:4n3)	0.9092	2.0629
	eicosapentaenoate (EPA; 20:5n3)	0.9697	1.6829
	docosapentaenoate (n3 DPA; 22:5n3)	1.0019	2.1902
	docosahexaenoate (DHA; 22:6n3)	1.0062	1.4737
	linolenate [alpha or gamma; (18:3n3 or 6)]	0.9729	1.8151
	docosapentaenoate (n6 DPA; 22:5n6)	0.8102	1.527
	dihomo-linoleate (20:2n6)	0.8467	1.6506
	mead acid (20:3n9)	0.6505	2.1117
Fatty acid Metabolism	myristoylcarnitine	0.929	1.8567
	palmitoylcarnitine	0.8664	1.5805
	stearoylcarnitine	1.0211	2.2988
	oleoylcarnitine	1.0648	1.5524
	myristoleoylcarnitine*	1.0617	1.6484
Fatty Acid, Monohydroxy	2-hydroxyoctanoate	0.7521	1.6325
	2-hydroxydecanoate	0.6441	2.0658
	3-hydroxyoctanoate	1.0549	1.9832
	3-hydroxydecanoate	1.1702	2.1847
	3-hydroxylaurate	1.2756	1.9255
	3-hydroxymyristate	1.072	1.6838
	5-hydroxyhexanoate	1.624	0.9847
	13-HODE; 9-HODE		

Alteration of RSV-induced DC activation by plasma from *L. johnsonii* supplemented mice



- BMDC were pre-incubated with **plasma** from supplemented animals at day 2 of RSV infection.
- DC were infected with RSV for 24 hrs and assessed for cytokine expression.
- Similar to the response of BMDC from supplemented mice, the plasma from *L. johnsonii* supplemented mice induced higher cytokine production.

Environment, Microbiome, Metabolic activity, and immunity





MAAP Collaboration

NIH, NIAID- P01AI089473



Christine Johnson, PhD, MPH
Ed Zoratti, MD
Kevin Bobbitt, PhD
Kim Woodcroft, PhD
Genesa Wegienka, PhD
Susan Havstad, PhD
Al Levine, PhD
Andrea Cassidy, PhD
Kyra Jones
Haejin Kim, MD
Alex Sitarik, MS
Karen Broski, PhD

Dennis Ownby, MD

Susan Lynch, PhD
Homer Boushey, MD
Kei Fujimura, PhD
Marcus Rauch, PhD
Ariane Panzer
Kaitlyn Lucy, PhD

Tine Demoor, PhD
Aaron Berlin
Andrew Rasky
Sihyug Jang, PhD
Wendy Fonseca, PhD
Nick Lukacs, PhD